



INTERNATIONAL JOURNAL
OF PHARMA AND BIO SCIENCES

Internationally indexed journal

Indexed in Chemical Abstract Services (USA), Index coppernicus, Ulrichs Directory of Periodicals, Google scholar, PSOAR, EBSCO, Open J gate, Proquest, EMBASE, etc.



Rapid and Easy Publishing

The “International Journal of Pharma and Bio Sciences” (IJPBS) is an international journal in English published quartely. The aim of IJPBS is to publish peer reviewed research and review articles rapidly without delay in the developing field of pharmaceutical and biological sciences.

Indexed in Elsevier Bibliographic Database (EMBASE)
SCImago Journal Rank 0.329
Impact factor 6.268* (SJIF)



Pharmaceutical Sciences

- Pharmaceutics
- Novel drug delivery system
- Nanotechnology
- Pharmacology
- Pharmacognosy
- Analytical chemistry
- Pharmacy practice
- Pharmacogenomics
- Polymer sciences
- Biomaterial sciences
- Medicinal chemistry
- Natural chemistry
- Biotechnology
- Pharmacoinformatics
- Biopharmaceutics

Biological Sciences

- Biochemistry
- Biotechnology
- Bioinformatics
- Cell biology
- Microbiology
- Molecular biology
- Neurobiology
- Cytology
- Pathology
- Immunobiology

ELSEVIER INDEXED JOURNAL

SNIP VALUE - 0.538

SJR - 0.274

CITESCORE - 0.35

SNIP - Source Normalised Impact per Paper

SJR - SCImago Journal rank

*Source - www.journalmetrics.com
(powered by scopus (ELSEVIER))*

ELSEVIER
EMBASE

ISSN 0975-6299

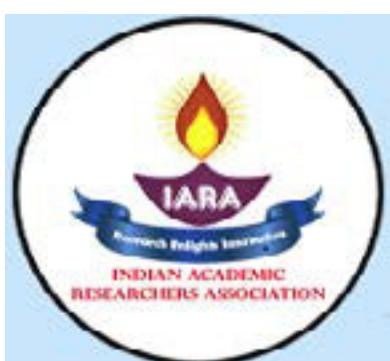
CODEN IJPBJ2

 **Crossref**

DOI: <https://doi.org/10.22376/ijpbs>

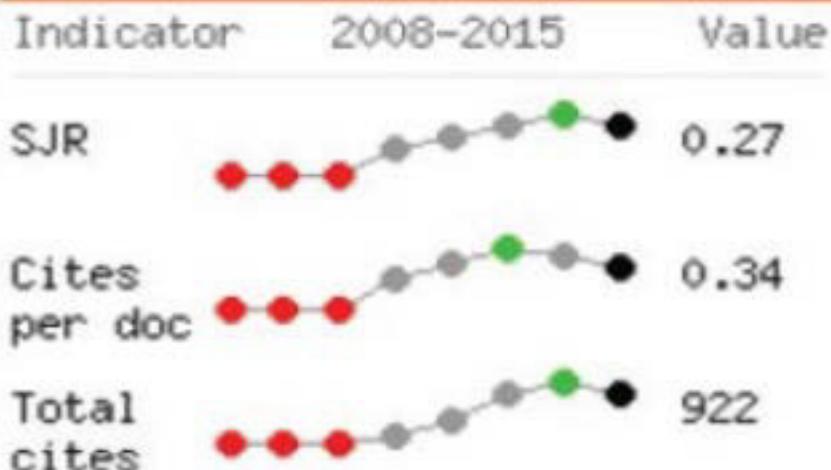
Google
Scholar
BETA

 **CrossMark**
← click for updates



 **INTERNATIONAL**
Scientific Indexing

International Journal of Pharma and Bio Sciences



www.scimagojr.com



A division of the American Chemical Society



CAS Source Index (CASSI) Search Tool



Arbeitsgemeinschaft
der deutschen
Familienorganisationen e.V.



Site designed and hosted by [Annals of Internal Medicine](#) / [American College of Physicians](#)



WZB

Berlin Social Science Center





DEVELOPMENT OF A SIMPLE, RAPID AND SPECIFIC RP-HPLC METHOD FOR THE ESTIMATION OF PYRIDOXINE HYDROCHLORIDE AND DOXYLAMINE SUCCINATE IN BULK AND COMBINED PHARMACEUTICAL DOSAGE FORMS

AMAR KUMAR KASTURI¹, KANAKA RAJU MEDICHERLA², N. BALAKRISHNAN^{3*}

¹*Department of Chemistry, Pacific University, Udaipur, Rajasthan-313003, India.*

²*Department of Human Genetics, Andhra University, Visakhapatnam, Andhra Pradesh-530 003, India.*

³*Principal Cum Professor, S A Raja Pharmacy College, Vadakkangulam-627116, Tamilnadu India.*

ABSTRACT

A rapid and specific method was developed and validated for the quantitation of Pyridoxine Hydrochloride and Doxylamine in bulk and combined pharmaceutical dosage forms. To achieve the chromatographic separation Shimadzu system has been used, equipped with LC-20AD high-performance liquid chromatography, degasser PGU-20A 5, variable wavelength programmable diode array detector SPD-M20A, auto sampler SIL-20 AC HT, and column oven CTO-10 A5 VP hic separation was achieved by using Shimadzu prominence LC-20AD high-performance liquid chromatography, equipped with degasser PGU-20A 5, variable wavelength programmable diode array detector SPD-M20A, auto sampler SIL-20 AC HT, and column oven CTO-10 A5 VP. XBridge C18, 4.6 mm X150 mm, 5 microns Make: Waters, Water's Corporation. Inc. The proposed method was validated for selectivity, precision, linearity and accuracy. The developed method was successfully applied to estimate the amount of Pyridoxine Hydrochloride and Doxylamine in bulk and combined dosage forms.

KEYWORDS: *RP-HPLC, Pyridoxine Hydrochloride (PYH), Doxylamine Succinate (DXA).*



N. BALAKRISHNAN*

**Principal Cum Professor, S A Raja Pharmacy College,
Vadakkangulam-627116, Tamilnadu, India.**

Received on: 11-06-2018

Revised and Accepted on: 02-08-2018

DOI: <http://dx.doi.org/10.22376/ijpbs.2018.9.4.p21-28>



[Creative commons version 4.0](#)

INTRODUCTION

Pyridoxine hydrochloride (PYH)¹ is chemically 3, 4-pyridinediacetonitrile, 5-hydroxy-6-methyl, hydrochloride (Figure 1). It is a water-soluble vitamin and involved principally in amino acid, carbohydrate, and fat metabolism². It is also required for the formation of haemoglobin.^{3,4,5} Doxylamine is a first-generation antihistamine.^{4,5} It can be used by itself as a short-term sedative and in combination with other drugs to provide night-time allergy and cold relief. Doxylamine is also used in combination with the analgesics paracetamol (acetaminophen) and codeine as an analgesic/镇静药 preparation, and is prescribed in combination with

vitamin B6 (pyridoxine) to prevent morning sickness in pregnant women. HPLC methods are useful in the determination of drugs in pharmaceutical formulations especially with those containing more than one active components. Therefore, the aim of this work was to develop a relatively simple HPLC method for simultaneous quantification of Pyridoxine hydrochloride (PYH) and Doxylamine (DXA). This paper describes the development and validation of reliable, simple, stable and economic reverse phase HPLC assay suitable for quality control in pharmaceutical industry due to its sensitivity, simplicity, selectivity and lack of excipients interference.

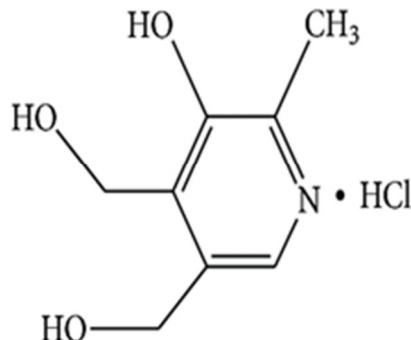


Figure 1
Chemical structure of Pyridoxine HCl

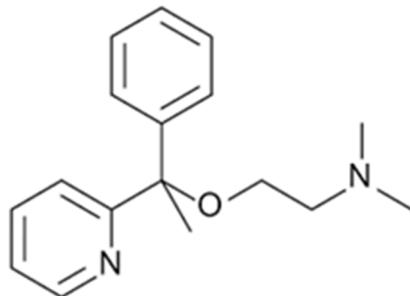


Figure 2
Chemical structure of Doxylamine

MATERIALS AND METHODS

Pure standards of PYH and DXA were obtained from Sigma-Aldrich Co.

Instrument and Chromatographic Condition

To achieve the chromatographic separation Shimadzu system has been used, equipped with LC-20AD high-performance liquid chromatography, degasser PGU-20A 5, variable wavelength programmable diode array detector SPD-M20A, auto sampler SIL-20 AC HT, and column oven CTO-10 A5 VP. XBridge C18, 4.6X150 mm, Make: Water's Corporation. Inc. The column temperature was kept at 30° C, and the mobile phase flow rate was maintained at 1.0 mL/min. The detector was Setat 210 nm.⁶ The injection volume was 10 µL, and the run time was 20 min for each injection. Other instruments such as pH meter, electronic weighing balance, and ultrasonic bath were also used.

Diluent Preparation

Diluent – 1
250 mg of sodium hydroxide in 1000 mL water

Diluent – 2
2 mL of orthophosphoric acid in 1000 mL of water

Standard stock preparation

PYH and DXA were weighed (25 mg each) and transferred to two separate 50 ml volumetric flasks and about 5 mL of diluent-1 and 30 ml of diluent -2 was added and sonicated to dissolve. Volumes were made up to the mark with diluent-2. Aliquot from the stock solution of PYH was appropriately diluted with mobile phase to obtain working standard of 100 $\mu\text{g mL}^{-1}$ of PYH and same way for DXA.

Assay sample preparation

Weigh accurately and transfer 10 numbers of Doxylamine Succinate and Pyridoxine Hydrochloride delayed release tablets into a 200 mL volumetric flask. Add 20 mL of diluent-1 and sonicate with intermittent shaking until to disintegrate. Add 120 mL of diluent-2 and sonicate for 15 minutes with intermittent shaking. Shake mechanically for 15 minutes. Dilute to volume with diluent-2 and mix well. Centrifuge a portion of the

Gradient programme**Composition of solvent for gradient programme**

| Time (min) | % of Mobile Phase A | % of Mobile Phase B |
|---------------|------------------------|------------------------|
| 0 | 100 | 0 |
| 12 | 40 | 60 |
| 14 | 100 | 0 |
| 20 | 100 | 0 |

Method validation

The method was validated in terms of the following parameters, system suitability, linearity, LOD, LOQ, specificity, accuracy, precision, robustness and ruggedness as per the ICH guidelines.

System Suitability

The chromatographic conditions were set as per the optimized parameters and steady baseline. Six replicates of working standard solution are injected and the Chromatograms are the mobile phase was allowed to equilibrate with stationary phase as was indicated by the recorded. The % relative standard deviation (%RSD).⁸ of retention time, asymmetry, theoretical plate count and peak areas were determined and the results were shown in Table 1.

Linearity

Accurately measured volume of the standard stock solution was diluted with diluents to get the final concentrations of Standard PYH as 5-25 $\mu\text{g/ml}$ and Doxylamine succinate standard as 20-100 $\mu\text{g/ml}$ respectively. Six different concentrations of the mixed standard drugs of Pyridoxine HCL and DXA were prepared for linearity studies and injected into the system (n=6). The response was measured as peak areas. Each concentration was prepared from individual stock solution. The peak areas were plotted versus concentrations to get the calibration curve.

Detection limit and quantification limit [LOD and LOQ]

The sensitivity of the simultaneous method of Pyridoxine HCL and Doxylamine succinate is estimated in terms of Limit of Detection (LOD) and Limit of Quantitation

above solution at 3000 rpm for 10 minutes. Dilute 5 mL of the above supernatant solution to 25 mL with diluent-2 and mix well. Filter through 0.45 μm PVDF filter by discarding the first 4 mL of the filtrate. (Concentration of about 100 $\mu\text{g/mL}$ of Doxylamine Succinate and Pyridoxine Hydrochloride).^{6,7}

Optimization of chromatographic condition

Mode of operation: Gradient

Column: XBridge C18, 4.6 mm X 150 mm, 5.0 μm .

Make: Waters Corporation.Inc ,

Flow rate: 1.0 ml/min

Column temperature: 30 $^{\circ}\text{C}$

Sample port temperature: 25 $^{\circ}\text{C}$

Injection volume: 10 μL

Run time: 20 minutes

Retention times: 4.6 minutes and 11.5 minutes for PYH and DXA respectively.

(LOQ). The LOD and LOQ⁹ were calculated using the formula.

$$\text{LOD} = 3.3 \times \sigma/S$$

$$\text{LOQ} = 10 \times \sigma/S$$

σ = Standard Deviation of Response

S = Slope of the calibration curve

The results were shown in Table.1 standard

Specificity

The specificity of the method was performed by injecting a blank solution (without any sample) and then a drug solution of 10 μl injected into the column, under optimized chromatographic conditions, to demonstrate the separation of both Pyridoxine hcl and Doxylamine.

Accuracy

The accuracy was carried out by adding known amounts of each analyte corresponding to three concentration levels (80%, 100%, 120%)¹¹⁻¹³ of the label claim to the excipients. At each level, six determinations were determined and the results are expressed as a percentage. Analyte recovered by the proposed method. The results are given in the Table.3.

Precision

The precision of the method was assessed by studying intra-day and inter-day variation. In the intra-day studies, standard and sample solutions were analyzed in triplicate on the same day and % RSD was calculated. In case of inter-day studies, standard and sample solutions were analyzed in triplicate on three consecutive days and % RSD were calculated. The results are shown in the table.4

Robustness

Robustness is a measure of capacity of analytical methods to remain unaffected by small deliberate variation of the operating conditions. It was tested by changing the flow rate, temperature and wavelength by ± 2 nm. The results are shown in the Table .5

Ruggedness

The ruggedness of the method was analyzed in different days and different chemists to check for any changes in the chromatograph, % RSD for the retention time and the area was calculated. The results are shown in the Table.6.

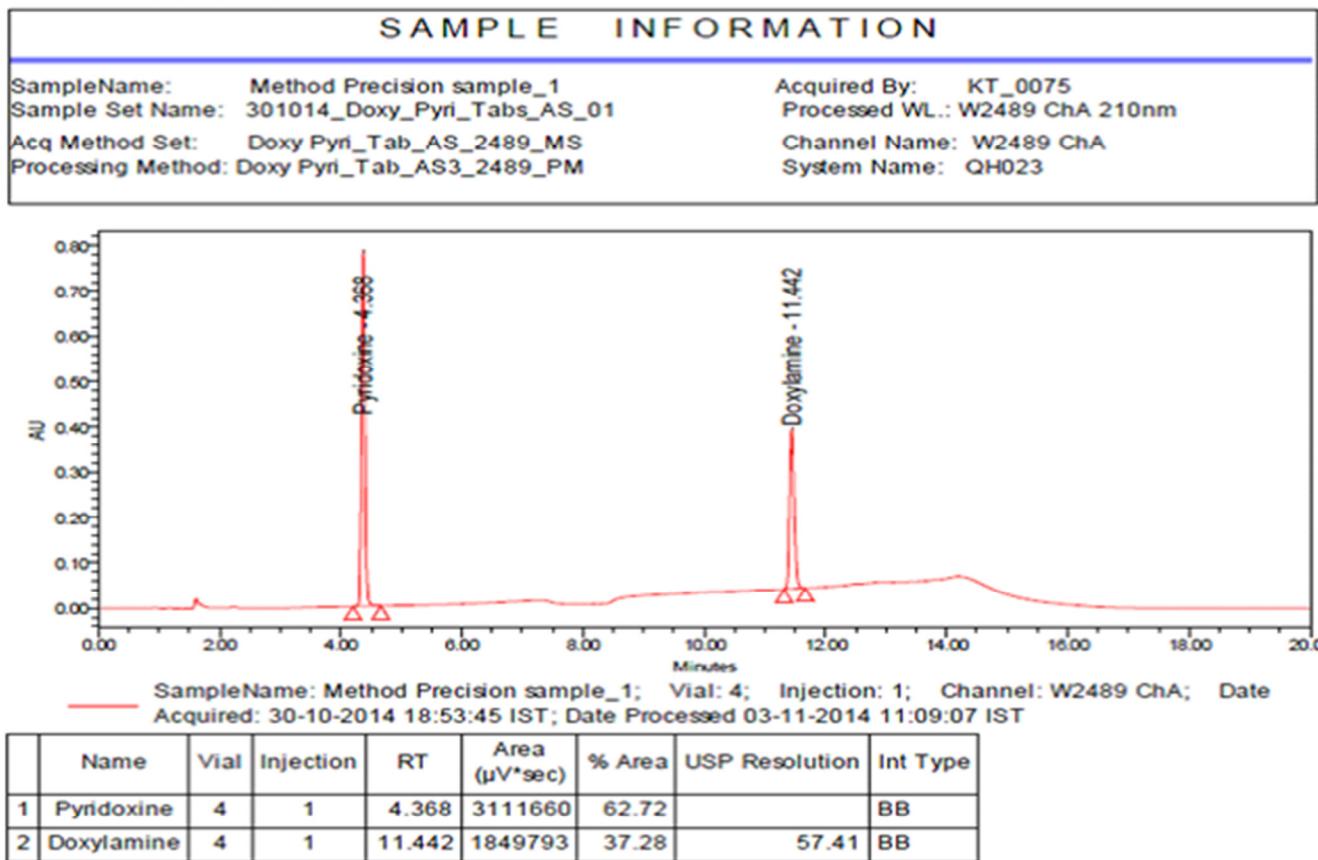
RESULTS AND DISCUSSIONS**Method Validation**

Figure 3
Optimised chromatogram of Pyridoxine Hcl and andDoxylamine.

System Suitability

The relative standard deviation (RSD) of Doxylamine and Pyridoxine peak area is NMT 2.0% from standard preparation. The USP plate and Pyridoxine peak is NLT

counts for Doxylamine 5000 from standard preparation. The tailing factor for Doxylamine and Pyridoxine peak is NMT 2.0 from standard preparation.

Table 1
System suitability.

| Injection No | Peak area for Doxylamine | Peak area for Pyridoxine |
|--------------------|--------------------------|--------------------------|
| 1 | 1896503 | 3185563 |
| 2 | 1893412 | 3160400 |
| 3 | 1854186 | 3106135 |
| 4 | 1869798 | 3142209 |
| 5 | 1864999 | 3138109 |
| Mean | 1875779 | 3146483 |
| % RSD | 0.98 | 0.93 |
| USP Tailing factor | 1.18 | 1.06 |
| USP Plate count | 107964 | 28327 |

Linearity Determination

Solutions of DXA and PYH at concentration levels from about 50% to 200% of standard solution were injected into HPLC system. The linearity graph was plotted from 50% to 200%. Six injections were performed at 50% level and at 200% level.¹⁴⁻¹⁶

RESULTS

Linearity and range of assay method is established by injecting series solutions of Doxylamine Succinate and Pyridoxine Hydrochloride. The data is shown in Table 2 and 3.

Table 2
Linearity study for Doxylamine Succinate.

| Sample No. | % level | Concentration ($\mu\text{g/mL}$) | Mean Peak Area |
|------------|---------|------------------------------------|----------------|
| 1 | 50 | 51.344 | 965375 |
| 2 | 75 | 77.017 | 1437913 |
| 3 | 80 | 82.151 | 1527854 |
| 4 | 100 | 102.689 | 1938429 |
| 5 | 120 | 123.227 | 2298332 |
| 6 | 150 | 154.034 | 2938521 |
| 7 | 200 | 205.379 | 3854584 |

Table 3
Linearity study for Pyridoxine HCL.

| Sample No. | % level | Concentration ($\mu\text{g/mL}$) | Mean Peak Area |
|------------|---------|------------------------------------|----------------|
| 1 | 50 | 51.192 | 1632788 |
| 2 | 75 | 76.788 | 2431554 |
| 3 | 80 | 81.907 | 2585474 |
| 4 | 100 | 102.384 | 3274215 |
| 5 | 120 | 122.861 | 3868837 |
| 6 | 150 | 153.577 | 4928177 |
| 7 | 200 | 204.769 | 6419923 |

Table 4
Linearity Plot of Doxylamine Succinate.

| Linear Regression Analysis for Doxylamine Concentration in $\mu\text{g/mL}$ vs. Area | |
|--|-----------|
| Correlation Coefficient Square (r^2) | 0.9995 |
| Slope | 18905.870 |
| Y-Intercept | 12151.215 |

Table 5
Linearity Plot of Pyridoxine Hydrochloride.

| Linear Regression Analysis for Pyridoxine Concentration in $\mu\text{g/mL}$ vs. Area | |
|--|-----------|
| Correlation Coefficient Square (r^2) | 0.9994 |
| Slope | 31403.857 |
| Y-Intercept | 31813.946 |

Detection limit and quantification limit (LOD and LOQ)**Pyridoxine Hydrochloride**

LOQ: 0.143 $\mu\text{g/mL}$

LOD: 0.047 $\mu\text{g/mL}$

Doxylamine Succinate

LOQ: 0.255 $\mu\text{g/mL}$

LOD: 0.084 $\mu\text{g/mL}$

Specificity

The specificity of the RP-HPLC method was determined by the complete separation of Pyridoxine HCL ((PYH) and Doxylamine succinate (DXA)^{17,18}as shown in Figure

5. The peaks obtained for (PYH)and DXA were sharp and have a clear baseline separation.

Accuracy

Known amount of DXA and PYH were spiked with placebo for Doxylamine Succinate and Pyridoxine Hydrochloride Delayed Release Tablets, 10mg/ml,in order to produce recovery at 50%, 100 % and 200% levels of the DXA and PYH working concentration 100 $\mu\text{g/mL}$. Spiked assay samples were prepared in triplicate, injected in duplicate and the percentage recovery was calculated.

Table 6
Method accuracy study- Doxylamine Succinate.

| Sample No. | Theoretical (%) | Mean Peak Area | % Recovery | Mean (%) Recovery | % RSD |
|------------|-----------------|----------------|------------|-------------------|-------|
| 1 | 50 | 893103 | 98.42 | 97.99 | 0.44 |
| 2 | 50 | 883742 | 97.54 | | |
| 3 | 50 | 888112 | 98.01 | | |
| 1 | 100 | 1764581 | 97.80 | 97.76 | 0.32 |
| 2 | 100 | 1770794 | 98.07 | | |
| 3 | 100 | 1759898 | 97.43 | | |
| 1 | 200 | 3537017 | 97.27 | 98.28 | 0.89 |
| 2 | 200 | 3562222 | 98.71 | | |
| 3 | 200 | 3566293 | 98.86 | | |

Table 7
Method accuracy study- Pyridoxine Hydrochloride.

| Sample No. | Theoretical (%) | Mean Peak area | % Recovery | Mean (%) Recovery | % RSD |
|------------|-----------------|----------------|------------|-------------------|-------|
| 1 | 50 | 1515131 | 102.12 | 101.47 | 0.69 |
| 2 | 50 | 1508084 | 101.57 | | |
| 3 | 50 | 1515042 | 100.72 | | |
| 1 | 100 | 3014239 | 102.19 | 102.08 | 0.51 |
| 2 | 100 | 3025362 | 102.54 | | |
| 3 | 100 | 3001739 | 101.51 | | |
| 1 | 200 | 5891677 | 99.81 | 100.30 | 0.42 |
| 2 | 200 | 5931930 | 100.55 | | |
| 3 | 200 | 5930250 | 100.56 | | |

Precision

Precision of the assay method was determined by injecting, in duplicate, six individual sample solutions of Doxylamine Succinate and Pyridoxine Hydrochloride Delayed Release Tablets, 10mg/ml. The samples were prepared as per the method.

Table 8
Precision study Doxylamine Succinate and Pyridoxine Hydrochloride.

| Sample No | DoxylamineSuccinate | | Pyridoxine Hydrochloride | |
|-----------|---------------------|---------|--------------------------|---------|
| | Mean peak area | % assay | Mean peak area | % assay |
| 1 | 1849235 | 99.02 | 3111204 | 100.04 |
| 2 | 1842286 | 98.65 | 3103431 | 99.79 |
| 3 | 1824276 | 97.68 | 3103350 | 99.79 |
| 4 | 1906478 | 102.08 | 3165773 | 101.80 |
| 5 | 1899109 | 101.69 | 3200017 | 102.90 |
| 6 | 1870864 | 100.18 | 3176516 | 102.14 |
| Mean | NA | 99.88 | NA | 101.07 |
| %RSD | NA | 1.75 | NA | 1.35 |

Robustness

Standard solution was prepared and injected into the chromatographic system as per the conditions specified in the method. The same standard solution was re-injected by changing one parameter at a time, keeping

other parameters constant. A set of system suitability data was calculated for standards injected under altered method conditions and compared against the values generated under normal method conditions.

Method Parameters

Flow Rate (Normal flow is 1.0 mL/min)

Flow minus → 0.9 mL/min

Flow plus → 1.1 mL/min

Column Operating Temperature (Normal temperature is 30°C)

Temperature minus → 25°C

Temperature plus → 35°C

Table 9
Robustness study- Retention time of Doxylamine Succinate.

| Parameters | | Retention Time (min) | Mean Peak area (n=5) | %RSD | USP Tailing factor | USP Plate count |
|--------------------------|------------------|----------------------|----------------------|------|--------------------|-----------------|
| Normal Condition | 1.0 mL/min, 30°C | 11.367 | 1947150 | 0.13 | 1.19 | 108113 |
| Flow Rate Minus | 0.9 mL/min | 11.816 | 2139573 | 0.11 | 1.20 | 112874 |
| Flow Rate Plus | 1.1 mL/min | 11.216 | 1948289 | 0.06 | 1.19 | 107352 |
| Column Temperature Minus | 25°C | 11.520 | 1945738 | 0.13 | 1.19 | 108851 |
| Column Temperature Plus | 35°C | 11.220 | 1945016 | 0.14 | 1.04 | 110073 |

Table 10
Robustness study - Retention time of Pyridoxine Hydrochloride.

| Parameters | | Retention Time (min) | Mean Peak area (n=5) | %RSD | USP Tailing factor | USP Plate count |
|--------------------------|------------------|----------------------|----------------------|------|--------------------|-----------------|
| Normal Condition | 1.0 mL/min, 30°C | 4.289 | 3376835 | 0.08 | 1.05 | 23438 |
| Flow Rate Minus | 0.9 mL/min | 4.650 | 3737177 | 0.06 | 1.07 | 26331 |
| Flow Rate Plus | 1.1 mL/min | 4.168 | 3378083 | 0.08 | 1.04 | 22504 |
| Column Temperature Minus | 25°C | 4.398 | 3386147 | 0.09 | 1.05 | 24330 |
| Column Temperature Plus | 35°C | 4.171 | 3371221 | 0.13 | 1.20 | 22448 |

CONCLUSION

The proposed method was validated in accordance with ICH guidelines and all the results obtained with this method are all within the limits. This method can be suitable for routine analysis in laboratories. Therefore, this method is simple, accurate, rapid, reliable method for simultaneous estimation of Pyridoxine HCL and Doxylamine Succinate.

AUTHOR CONTRIBUTION STATEMENT

Conceived and designed the experiments: Amar

REFERENCES

1. Rajput S, Pathak A. Simultaneous derivative spectrophotometric analysis of doxylamine succinate, pyridoxine hydrochloride and folic acid in combined dosage forms. Indian J Pharm Sci. 2008 Aug 12;70(4):513-7. Available from: <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2792541/>
2. Melo L, Collins C, Jardim I. New materials for solid-phase extraction and multiclass high-performance liquid chromatographic analysis of pesticides in grapes. J Chromatogr A. 2004 Apr 01;1032(1-2):51-8.
3. Pacheco-Palencia L, Talcott S. Chemical stability of açai fruit (*Euterpe oleracea* Mart.) anthocyanins as influenced by naturally occurring and externally added polyphenolic cofactors in model systems. Food Chemistry. 2010 Jan 12;118(1):17-25.
4. K. D. Tripathi. Essentials of Medical Pharmacology. Jaypee Brother's Medical Publishers, New Delhi 5th ed. 2003;p. 821-3.
5. Sweetman S.C, Martindale Ed. The Complete drug reference, Pharmaceutical press; 2002;p.1219-20.
6. Argekar A, Sawant J. Simultaneous Determination of Pyridoxine Hydrochloride and Doxylamine Succinate from Tablets by Ion Pair Reversed-Phase High-Performance Liquid Chromatography (RP-HPLC). Drug Dev. Ind. Pharmacy. 1999;25(8):945-50.
7. K. Li. "Simultaneous determination of nicotinamide, pyridoxine hydrochloride, thiamine mononitrate and riboflavin in multivitamin with minerals tablets by reversed-phase ion-pair high performance liquid chromatography. Biomed Chromatogr. 2002;16(8):504-7. Available from: http://quimica.udea.edu.co/~carlopez/cromatohplc/vitaminas_rp_par_ionico.pdf
8. M. Alagar Raja, M. Samatha, B. David et al. Analytical method development and validation of acetaminophen, dextromethorphan hydro bromide doxylamine succinate in soft gel capsule dosage

Kumar Kasturi and N.Balakrishnan. Performed the experiments: Amar Kumar Kasturi. Analyzed the data: Amar Kumar Kasturi, Kanaka raju Medicherla and N. Balakrishnan. Contributed reagents/materials/analysis tools: N. Balakrishnan. Wrote the paper: Amar Kumar Kasturi and Kanaka raju Medicherla.

CONFLICT OF INTEREST

Conflict of interest declared none.

form by using RP-HPLC. *World J Pharm Pharm Sci.* 2013;2(6):5852–62.

9. El-Gindy A. Spectrophotometric and LC determination of two binary mixtures containing Pyridoxine hydrochloride. *J. Pharm. Biomed. Anal.* 2003;32(2):277-86.

10. Nunes B, Cruz A, Faria J, Sant' Ana A, Silva R, Moura M. A survey on the sanitary condition of commercial foods of plant origin sold in Brazil. *Food Control.* 2010;21(1):50-4.

11. Islam S, Hossain M, Nahar N, Mosihuzzaman M, Mamun M. Application of High Performance Liquid Chromatography to the Analysis of Pesticide Residues in Eggplants. *J. Appl. Sci.* 2009 Nov 12;9(5):973-7.

12. P. Jin, L. Xia, Z. Li, N. Che, D. Zou, and X. Hu. Rapid determination of thiamine, riboflavin, niacinamide, pantothenic acid, pyridoxine, folic acid and ascorbic acid in Vitamins with Minerals Tablets by high-performance liquid chromatography with diode array detector. *J. Pharm. Biomed. Anal.* 2012;70:151–7.

13. P. C. P. Rosa and I. C. S. F. Jardim. Simultaneous determination of clobutinol hydrochloride and doxylamine succinate from syrups by RP HPLC using a new stationary phase containing embedded urea polar groups. *Braz. J. Pharm. Sci.* 2012;48(2):315–23.

14. M. D. Saddam Nawaz. A new validated stability indicating RP-HPLC method for simultaneous estimation of pyridoxine hydrochloride and meclizine hydrochloride in pharmaceutical solid dosage form. *Chromatogr. Res. Int.* 2013;Article ID 747060:1- 7. Available from: <https://www.hindawi.com/journals/cr/2013/747060/>

15. K. Reema, V. Itishree, and N. Shantaram, Jagdish. Method development and validation for the simultaneous estimation of B-group vitamins and atorvastatin in pharmaceutical solid dosage form by RP-HPLC. *Int. J. Pharm. Chem. Biol. Sci.* 2013;3(2):330–5.

16. S. K. Dhal and R. Sharma. Development and validation of RP-HPLC method for simultaneous determination of pyridoxine hydrochloride, isoniazid, pyrazinamide and rifampicin in pharmaceutical formulation. *Chemia Analityczna.* 2009;54(6):1487–1500.

17. H. Soni, A. K. Singhai, K. Mishra, and S. Sharma. Simultaneous determination of vitamins B1, B2 and B6 in multivitamin tablet and biological fluid by RP-HPLC. *Int. J. Pharm. Sci. Res.* 2012;3(7):2163–7.

18. N. Yantih, D. Widowati, Wartini, and T. Aryani. Validation of HPLC method for determination of Thiamine hydrochloride, Riboflavin, Nicotinamide, and Pyridoxine hydrochloride in syrup preparation. *Canadian Journal on Scientific and Industrial Research.* 2011;2 (7):269–77.

International Journal of Pharma and Bio Sciences is also cataloged in the following universities libraries

United States Universities



UC Berkeley Libraries,
California, United States



UNIVERSITY *of* WASHINGTON

University of Washington Libraries,
Seattle, Washington, United States



Branford Price Millar Library,
Oregon, United States



THE OHIO STATE
UNIVERSITY
UNIVERSITY LIBRARIES

Colombus, Ohio, United States



Tucson, Arizona, United States



Houston Cole Library
Alabama, United States



Miami, Florida, United States



Ralph Brown Draughon Library,
Auburn, United States



Oklahoma, United States

United Kingdom Universities



UNIVERSITY OF
BIRMINGHAM

Birmingham University Library,
Birmingham , United Kingdom



CENTRAL CONNECTICUT STATE UNIVERSITY
ELIHU BURRITT LIBRARY

New Britain, Connecticut



Bodleian Libraries,
Oxford, United Kingdom

Library

Lancaster
University



Lancaster, England



University of
BRISTOL | Library

Bristol, England

Other Universities



James Cook University Library,
Singapore



The university of Manchester Library,
Singapore



NUS
National University
of Singapore

NUS
Libraries



University
of Glasgow



THE
UNIVERSITY OF
BRITISH
COLUMBIA

UBC Libraries,
Vancouver, Canada



LUND
UNIVERSITY

Lund University Libraries,
Sweden

EBERHARD KARLS
UNIVERSITÄT
TÜBINGEN

Tübingen, Germany



UNIVERSITY
LIBRARY



Heidelberg University,
Germany



Geneva Foundation for Medical Education and Research
Geneva - Switzerland

**And indexed /catalogued in many more indexing bodies
and universities**

For complete list of indexing kindly visit our journal webpage (www.ijpbs.net)

For any query / submit manuscript /feedback kindly visit www.ijpbs.net

Authors/readers/users may visit our journal web page (home page and instruction to authors) for any further details



Submit your manuscript/feedback/queries to “contact” of
www.ijpbs.net